

## Learning Objectives

- Why perform cytology
- What is required
- Collection techniques
- Slide preparation and optimizing staining
- Slide examination including normal skin and artifacts
- Examination of aspirates of solid tissues and masses

## Why Perform Cutaneous Cytology?

- Most common diagnostic test performed in veterinary dermatology practice
- Provides the basis for empirical selection of antimicrobial drugs
- Adds diagnostic etiologic information based on cell type or structures present → medical vs. surgical management
- Best to correlate with results of culture and biopsy

## Why Perform Cutaneous Cytology?

- May be superior to histopathology for some microorganisms
- Safe for patient
- Cost effective
- Easily self taught
- Good business practice builder while improving the quality of medicine offered

## What Cytology Can't Do

- Determine tissue architecture
  - Precise diagnosis of neoplasm type not always possible (i.e. spindle cell)
  - Assess invasiveness
  - Assess distribution
- If poorly cellular
  - May not be representative
  - Negative results do not rule out disease

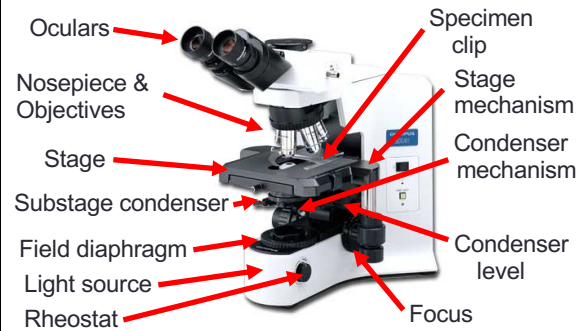
## What Cytology Can't Do

- Grade mast cell tumors
- Predict malignant/benign behavior of some neoplasms: mammary, melanoma, endocrine, hepatocellular
- Distinguish infiltrative from encapsulated lipoma
- Diagnose small cell variety lymphoma (sometimes)
- Distinguish granulation tissue from sarcoma (sometimes)

### What Do You Need?

- Binocular microscope with strong light source and high quality lenses
  - Ideal: 4x, 10x, 40x and 100x (oil immersion)
- Slides (frosted edge preferred)
- Immersion oil
  - Usually Type A or B
- Stain - *Diff-Quik*®
  - Quick and easy Romanowsky-type stain

### Anatomy of the Light Microscope

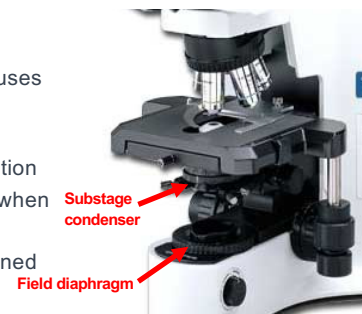


### Microscope Maintenance

- Annually – professional cleaning, service and lubrication
- Daily – dust cover
- As needed – ocular and lens cleaning
  - LENS paper ONLY
  - Alcohol based cleaners
  - Xylene (CAUTION with plastic lenses and some lens sealants)

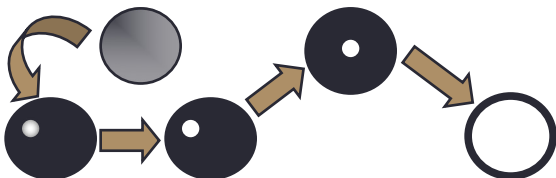
### Kohler Illumination

- AKA “critical focusing”
- Centers and focuses light onto the specimen for maximum resolution
- Best to perform when switching from unstained to stained specimens



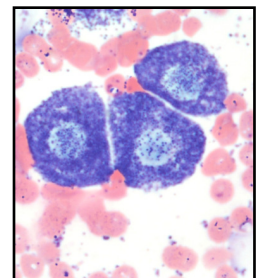
### Steps to Kohler Illumination

1. With slide on the stage, focus at 10x
2. Open substage condenser diaphragm fully
3. Close field diaphragm until only a dot of light is visible – FOCUS and CENTER
4. Re-open field diaphragm maximally



### Collection Techniques Impression Smear

- Identify inflammatory and neoplastic cells, as well as organisms that will aid in diagnosis
- Touch slide directly to lesion if moist or exudative

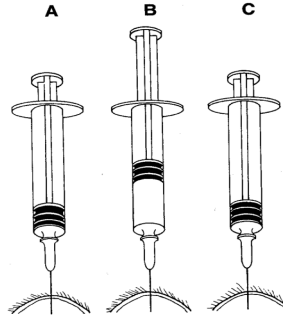


## Fine Needle Aspiration

- Indications: nodules and plaques
- Identify inflammatory and neoplastic cells as well as infectious organisms
- Avoid zones of inflammation and/or necrosis for best specimen
- Technique: aspiration vs. needle "core biopsy"
  - Aspiration: 6 ml syringe with 20-22 gauge needle
  - Insert needle into lesion



- Aspirate about 1 ml vacuum
- Move needle within lesion → repeat aspiration
- Release vacuum, extract syringe from lesion

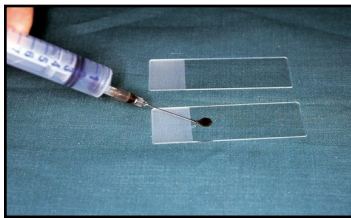


(A) insert  
(B) aspirate  
(C) release  
(repeat)

Cowell and Tyler's Diagnostic  
Cytology and Hematology of the  
Dog and Cat, 4th edition

## Fine Needle Aspiration

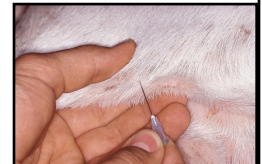
- Remove needle and add ~2 cc of air into the syringe
- Replace needle and push contents onto a clean glass slide



## Fine Needle Aspiration

### Needle core technique

- Insert needle into lesion
- Rotate needle on axis and redirect while within the lesion
- Repeat insertion
- Add ~2 cc of air to a 3 or 6 cc syringe
- Replace needle and push contents onto a clean glass slide



## Slide Preparation

- Collected materials are allowed to dry on slide
- Oily (adipocytes), waxy or dry samples may be heat fixed
- Diff-Quik®
  - ✓ Minimum of five 1 second dips in each stain
  - ✓ Rinse with water
  - ✓ Air dry or use hair drier
  - ✓ Gently blot with bibulous paper



## Slide Preparation

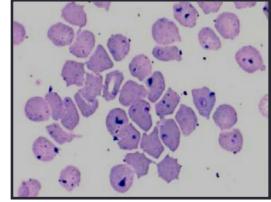
- Staining duration depends on specimen thickness
- Diff-Quik®
  - Fast, high contrast stain
  - Lacks cytoplasmic and nuclear detail
  - May miss cytoplasmic granules
- New methylene blue
  - Wet or unfixed slide may be stained
  - Little advantage over Diff-Quik®

## Getting the Most from DiffQuik®

- Storage
- “Dirty” vs. “Clean” specimens
- Replace stain at least once weekly
- Adapting staining to specimen thickness
- Avoid formalin exposure
- Stain soon after specimen has dried thoroughly

## Top Causes of Poor Staining

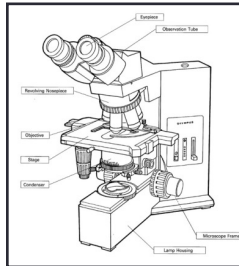
- Stain precipitate
- Stain contaminants
- Inappropriate specimen drying
- Too little fixative time
- Too much stain time
- Exposure to formalin (fumes or liquid)
- Prolonged delay before staining



## Slide Examination

- Stained specimens
- Cytology and hematology
- Stain provides contrast
- Settings for optimal light

**ESTABLISH  
KOHLE  
ILLUMINATION**

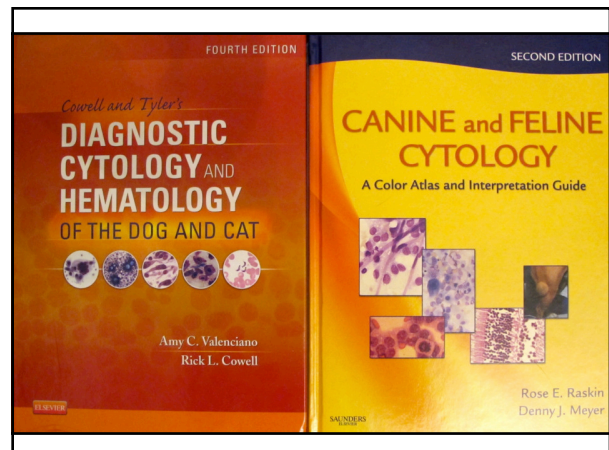


## Why is the 40x Blurry?

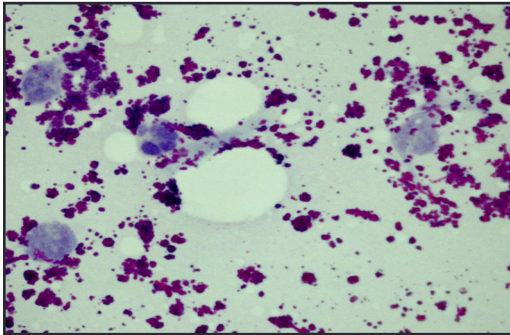
- Designed for use with cover slipped specimens
- HOW TO WORK AROUND:
  1. Stain and dry specimen as usual
  2. On top of stained specimen, place 1-2 drops immersion oil
  3. On top of oil, gently place cover slip (avoid bubbles!)
  4. 40x resolution should be improved
  5. Oil can be wiped off *gently* with Kleenex tissue

## Slide Examination

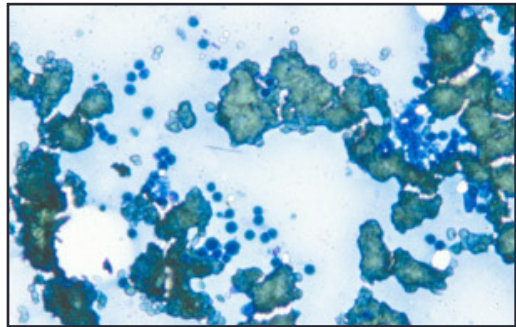
- Practice learning what is normal
- Learn to recognize artifacts (stain precipitate, hair, cotton fibers, pollen) that occur with cutaneous sampling
- Consider evaluating one sample in house and sending the other to a clinical pathologist
- PRACTICE!



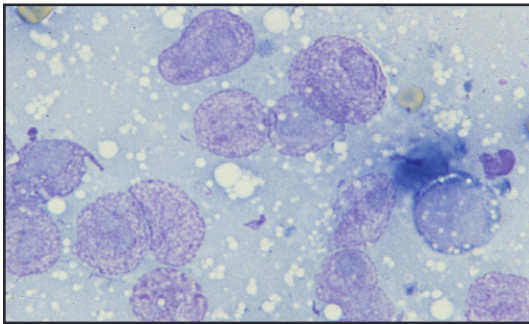
### Artifacts – Aqueous Lubricant



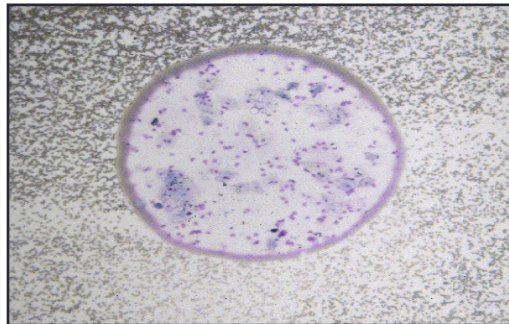
### Artifacts – Inadequate Spreading



### Artifacts – Cell Rupture



### Artifacts – Contaminants

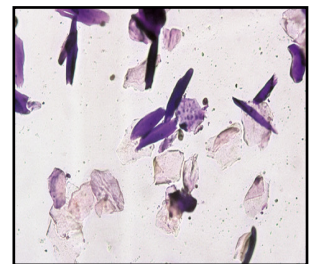


### Normal vs. Abnormal

- Normal skin
  - < 1 organism (yeast, cocci, rod)/OIF
  - No inflammatory cells
- Ears
  - Malassezia
    - ✓ Cats: > 1 Malassezia/OIF → significant
    - ✓ Dogs: > 3 Malassezia/OIF → significant
  - Cocci
    - ✓ >5 cocci/OIF → significant
    - ✓ >1 rod/OIF → significant

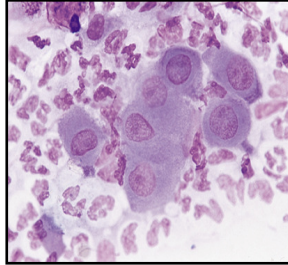
### Cytology of Normal Skin

- Epidermis
  - Corneocytes
    - Anucleated
    - Flat
    - Often folded



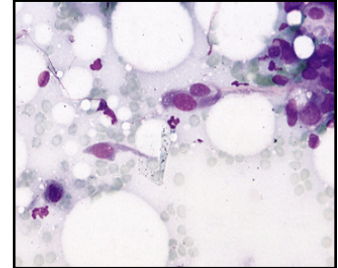
### Cytology of Normal Skin

- Epidermis: basal and spinous keratinocytes
  - Nucleated
  - Round nuclei
  - Basophilic cytoplasm



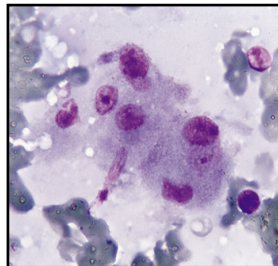
### Cytology of Normal Skin

- Dermis
- Fibroblasts
  - Spindle shaped
  - Oval nuclei



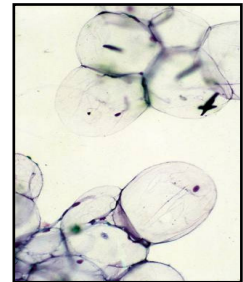
### Cytology of Normal Skin

- Dermis: sebocytes
  - Sebaceous glands
  - Round nucleus
  - Abundant cytoplasm with small clear vacuoles



### Cytology of Normal Skin

- Subcutis: adipocytes
  - Small displaced nucleus
  - Abundant clear cytoplasm (lipid)



## HOW TO EXAMINE ASPIRATES OF SOLID TISSUES AND MASSES

### Logical Approach to the Specimen

- Scan entire slide at low mag (4x and 10x)
  - Evaluate staining quality, cell density, arrangement of cells, hemodilution
  - Are cells intact or lysed?
  - Do cells "match" the site sampled?
  - Find an area to examine at higher magnification
- Examine areas at higher magnification (40x and 100x)
  - Identify normal cells
  - Identify inflammatory cells, infectious agents, neoplastic cells

## Common Questions to Answer

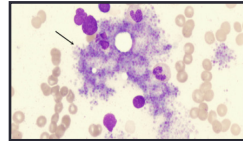
- Is there hemodilution or hemorrhage?
- Is there inflammation?
  - What kind of inflammation?
  - What is causing the inflammation?
  - Is there an infectious agent?
- Is there a neoplastic population?
  - What type of neoplasia (round, epithelial, spindle cell)?
  - Is it benign or malignant?

## Hemodilution or Hemorrhage?

Procedural blood contamination

Look for:

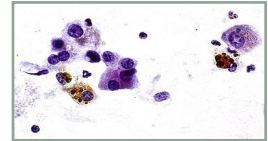
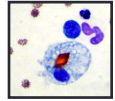
- Leukocytes in proportion to blood
- Platelet clumps



Hemorrhage

Look for:

- Macrophages consuming RBCs
- Hemosiderin
- Hematoidin



## Combinations – TRICKY!

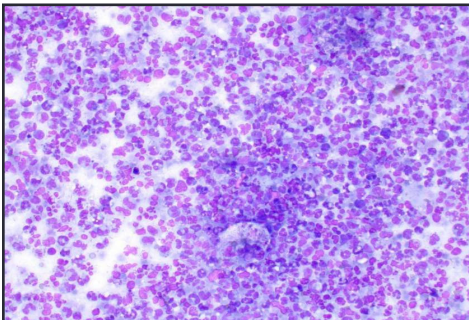
- Inflammation/infection + atypical cells = primary inflammatory lesion with dysplastic change OR
- Primary neoplastic lesion with superimposed inflammation, infection, hemorrhage and/or necrosis
- GET A BIOPSY!



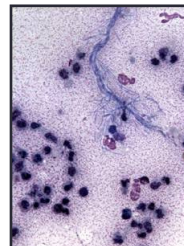
## If Inflammatory. . .

*Offers clue as to cause since different types of inflammation are seen with different etiologic agents*

## NEUTROPHILIC

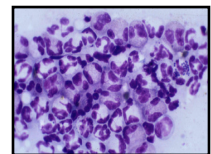


## NEUTROPHILIC



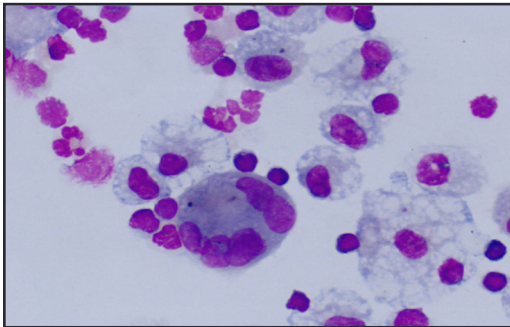
Feline abscess

- Trauma
- Foreign body
- Bacterial infection
- Immune-mediated disease
- Neoplasia



Deep pyoderma

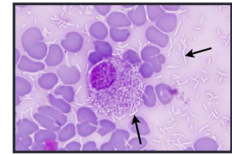
## GRANULOMATOUS



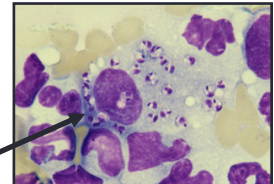
## GRANULOMATOUS

Pyogranulomatous =  
with neutrophils

- Mycobacteria
- Fungi
- Protozoa
- Foreign body
- Hemorrhage
- Neoplasia



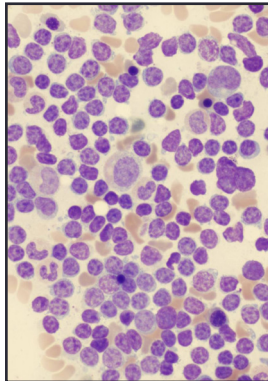
Mycobacterium



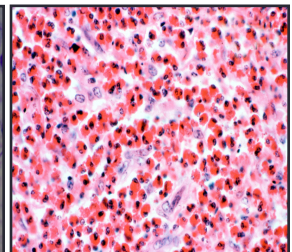
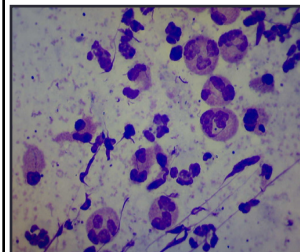
Canine Leishmania

## LYMPHOCYTIC

- Foreign body (vaccine granuloma)
- Immune-mediated disease
- Regressing histiocytoma
- Secondary to neoplasia (cell-mediated response)
- Lymphoma

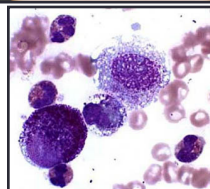
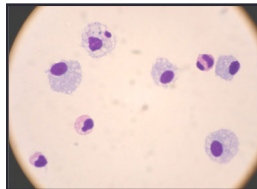


## EOSINOPHILIC



## EOSINOPHILIC

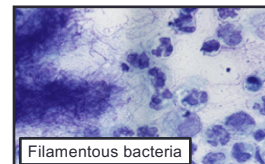
- Parasitic
- Fungal
- Hypersensitivity
- Foreign body
- Eosinophilic disorders
- Neoplasia



Canine MCT

## INFECTIOUS AGENTS

- Bacteria
- Fungi/algae
- Viral inclusions
- Protozoa
- Parasites



Filamentous bacteria



Fungal hyphae

## Neoplasia

For a diagnosis, must find:

- **Atypical** cells
- Cells in an **abnormal** location
- Cells in **abnormal** proportions

Inflammatory cells + atypical cells =  
*USE CAUTION*

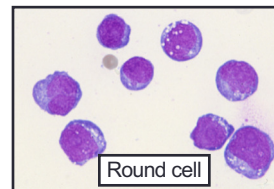
## Neoplasia - Steps in Evaluation

1. Determine cell type
2. Evaluate for benign vs. malignant

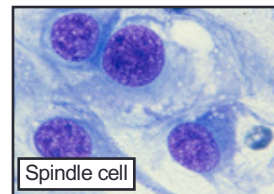


## Three Tumor Types

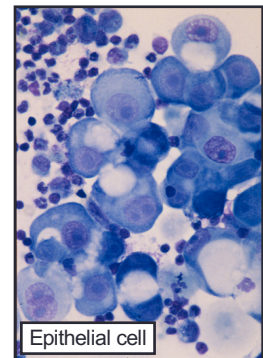
- **Round** - non-adherent, round-oval
- **Epithelial** - display cell-cell adhesion
- **Spindle (mesenchymal) cell** - non-adherent, fusiform



Round cell



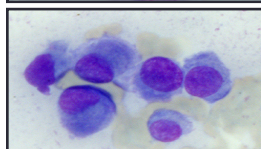
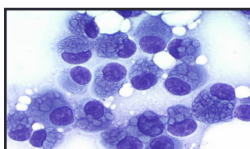
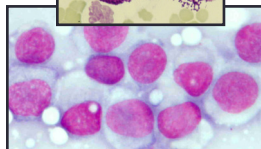
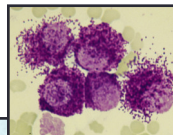
Spindle cell



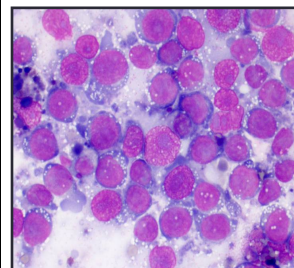
Epithelial cell

## Round Cell Tumors

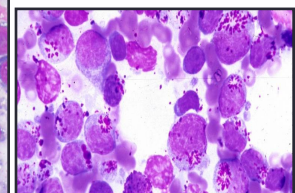
- Lymphoma
- Mast cell tumor
- Transmissible venereal tumor (TVT)
- Cutaneous histiocytoma
- Plasmacytoma



## Round Cell Tumors: Lymphoma



Vacuolated

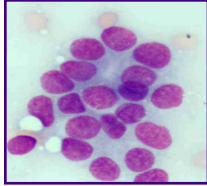


Granulated

## Epithelial Tumors – Generic Names

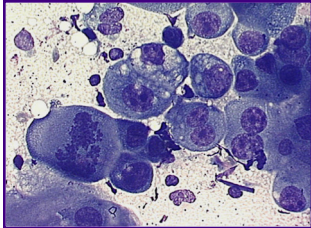
### BENIGN

- Epitheliomas
- Adenomas



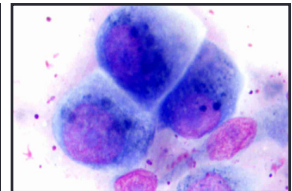
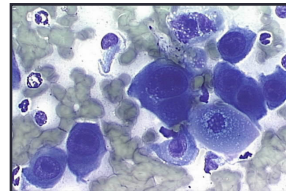
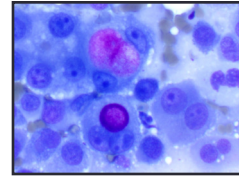
### MALIGNANT

- Carcinomas
- Adenocarcinomas

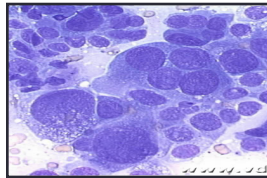
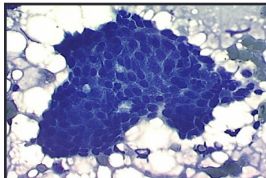
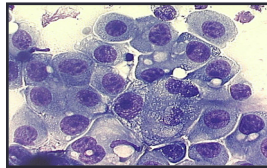
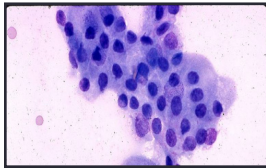


## Epithelial Tumors

- Trichoblastoma
- Transitional cell carcinoma
- Sebaceous adenoma
- Mammary adenocarcinoma

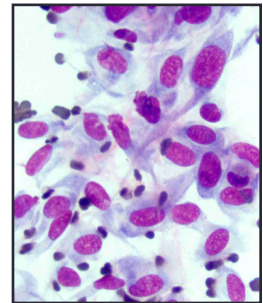


## Epithelial Tumors



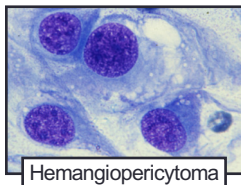
## Spindle Cell Tumors

- Reactive proliferation, benign and malignant neoplasms have considerable morphologic overlap
- BIOPSY OFTEN NEEDED*
- Benign = fibroma
- Malignant = sarcoma

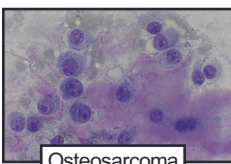


## Spindle Cell Tumors

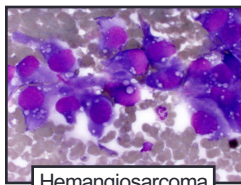
- Hemangiopericytoma
- Osteosarcoma
- Lipoma
- Chondrosarcoma
- Hemangiosarcoma



Hemangiopericytoma

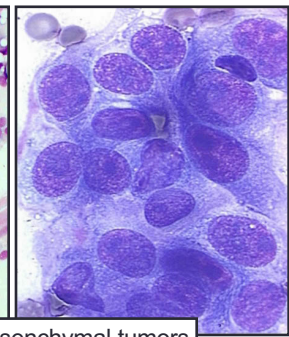
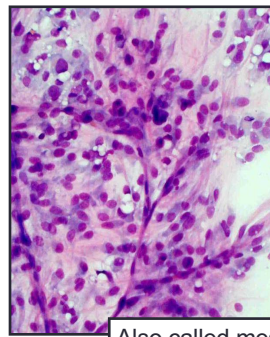


Osteosarcoma



Hemangiosarcoma

## Spindle Cell Tumors



Also called mesenchymal tumors



A diagram showing four epithelial cells. Each cell has a large, dark blue nucleus and a smaller, lighter blue nucleolus. The cells are connected by cell-cell junctions, represented by black lines. The cells are arranged in a cluster, with one cell at the top, one at the bottom, and two on the sides. The cell on the right is slightly separated from the others.

[illegible]

## Criteria of Malignancy

